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Low oxygen waters limited habitable space for early animals

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The oceans at the start of the Neoproterozoic Era (1000-541 million years ago, Ma) were dominantly anoxic, but may have become progressively oxygenated, coincident with the rise of animal life. However, the control that oxygen exerted on the development of early animal ecosystems remains unclear, as previous research has focussed only on the identification of fully anoxic or oxic conditions, rather than intermediate redox levels. Here, we report anomalous cerium enrichments preserved in carbonate rocks across bathymetric basin transects from nine localities of the Nama Group, Namibia (~550-541 Ma). In combination with Fe-based redox proxies, these data suggest that low oxygen conditions occurred in a narrow zone between well-oxygenated surface waters and fully anoxic deep waters. Although abundant in well-oxygenated environments, early skeletal animals did not occupy

oxygen impoverished regions of the shelf, demonstrating that oxygen availability (likely $>10\ \mu\text{M}$) was a key requirement for the development of early animal-based ecosystems.

Geochemical proxies based on Fe-S-C and trace metal systematics have been widely used to reconstruct the progressive oxygenation of the oceans during the Neoproterozoic and Cambrian¹⁻⁷. Accumulating evidence indicates that the deep oceans were dominantly anoxic and ferruginous (Fe-containing) throughout most of the Precambrian, with euxinic (sulphidic) mid-depth waters prevalent along continental margins from ~ 1.8 to 1.0 billion years ago (Ga)^{1,6,8}. From ~ 1.0 to 0.58 Ga, however, euxinic mid-depth waters became less prevalent and ferruginous conditions expanded, with oxic conditions still largely restricted to surface waters^{1,8,9}. The oxygenation of the deeper marine realm was both protracted and spatially heterogeneous, with some marine basins recording persistent deep-water oxygenation from ~ 580 Ma, whereas regional anoxia remained a feature of some deeper shelf environments into the Cambrian, ~ 520 Ma^{4,5,7,10} and beyond.

The course of Neoproterozoic oxygenation, and cause and effect associations with the appearance of animals, remains controversial^{4,11-13}. While modern soft-bodied sponge-grade animals may tolerate oxygen concentrations as low as $1.25\text{-}10\ \mu\text{M}$ ¹⁵, new innovations in the late Ediacaran, such as motility¹⁶, the rise of predation, and skeletonisation¹⁷⁻¹⁹, are all hypothesised to have required higher levels of oxygen²⁰. However, the oxygen demands of early animals are unconstrained, and observations

from modern biota cannot necessarily be applied to early animals of unknown affinity. Furthermore, while soft-bodied and skeletal Ediacaran fauna dominantly occur in sediments interpreted to have been deposited from oxic waters, fossil occurrences have also been reported in sediments characterised by anoxic geochemical signals^{5,14}. In the latter case, this may be because some early complex organisms were able to colonise habitats during fleeting periods of oxia (such short-lived oxygenation is difficult to detect by geochemical proxies that tend to integrate relatively long periods of time). In both of the above cases, however, there is uncertainty as to whether early animal evolution occurred under fully oxygenated conditions, or whether intermediate redox conditions were more prevalent, which by extension suggests that the oxygen requirements of more complex organisms were lower^{3,15}. An in-depth understanding of these links is currently hampered, however, by the inability of most redox proxies to distinguish between fully oxygenated and intermediate redox states, including nitrogenous or manganous conditions, which may overlap with low concentrations of oxygen^{16,17}. Indeed, it is possible that 'oxic' horizons identified through Fe and trace element geochemistry may in fact have formed under low oxygen conditions (but not fully anoxic), at levels insufficient to support diverse skeletal animal communities.

In oxic environments, Ce(III) is oxidised to insoluble Ce(IV) and preferentially scavenged relative to the rest of the rare earth elements and yttrium, REY¹⁸. The standard reduction potential of Ce(IV) (+1.61°V) is closer to Mn(IV) (+1.23°V) than Fe(III) (+0.77°V), and Ce oxidation is catalysed on the surface of Mn

(oxyhydr)oxides¹⁹. Therefore, the redox cycling of Ce in seawater is closely related to Mn(II)/Mn(IV) transformations, which occur at a higher redox potential than the Fe(II)/Fe(III) couple, and hence Mn cycling is more sensitive to intermediate redox conditions^{18–21}. Ce anomalies (Ce_{SN}/Ce_{SN}^*) are calculated here based on relative enrichments or depletions in shale-normalised Ce ($[Ce]_{SN}$) compared to neighbouring non-redox sensitive REY:



$$Ce_{SN}/Ce_{SN}^* = \frac{[Ce]_{SN}}{([Pr]_{SN})^2/[Nd]_{SN}} \quad (1)$$

Due to the accumulation of Ce(IV) on the surface of Mn (oxyhydr)oxides, oxic seawater becomes Ce-depleted and exhibits a negative Ce anomaly (<0.9)¹⁸ (see Methods and Supplementary Information for discussion of Ce/Ce* thresholds). These Mn (oxyhydr)oxides may be buried intact in sediments beneath oxic bottom waters, or may dissolve in the water column if they encounter low oxygen waters, releasing excess Ce. Therefore, waters beneath the Mn(IV)/Mn(II) redoxcline commonly exhibit a positive Ce anomaly (>1.3)^{20–22}. Positive Ce anomalies have been recorded alongside Mn enrichments in some modern waters, including Lake Vanda, Antarctica (Ce_{SN}/Ce_{SN}^* up to 2.3, Fig 1)²⁰, in anoxic brines in the eastern Mediterranean (Ce_{SN}/Ce_{SN}^* up to 2.43, Fig 1)²², and in the deep-marine Cariaco Basin (Ce_{SN}/Ce_{SN}^* up to 1.21)²¹.

Water column REY, and associated Ce anomalies, are **thought to be** preserved in **non-skeletal** carbonate rocks without fractionation²³. Carbonate-bound REY are **relatively** robust to diagenetic alteration²³ and dolomitisation²⁴, **but any alteration**

of the Ce anomaly can be identified using non-redox sensitive REY anomalies, such as the Y/Ho ratio, which would also be altered away from seawater patterns^{25,26}. We use a sequential dissolution method that targets REY in the carbonate phase, preventing contributions from sedimentary (oxyhydr)oxides or clays²⁶, which would carry a non-seawater signature. The majority of samples are very pure calcites with low siliciclastic components, but where samples are partially dolomitised they have been treated differently during leaching²⁶. The resulting REY data have been screened for traditional seawater features (Y/Ho ratios >67), and samples with evidence for diagenetic alteration or contributions from non-carbonate phases have been excluded from the presented Ce/Ce* data (see Methods and Supplementary Information).

We additionally utilise redox interpretations based on published Fe speciation data for these carbonate samples⁵. Fe-speciation distinguishes anoxic from oxic water column conditions through enrichments in highly reactive Fe (Fe_{HR}) relative to total Fe (Fe_{T})^{1,27}. Anoxic enrichments in Fe_{HR} occur due to the water column formation of either pyrite under euxinic conditions²⁷, or non-sulphidized Fe_{HR} minerals (such as Fe oxides or carbonates) under anoxic ferruginous conditions¹ (see Methods). Although originally calibrated for siliciclastics^{1,27}, enrichments in $\text{Fe}_{\text{HR}}/\text{Fe}_{\text{T}}$ can also be identified in carbonates deposited under anoxic water column conditions²⁸. These Fe_{HR} enrichments can far exceed Fe_{HR} contents expected under normal oxic deposition, where trace amounts (~0.1 wt%) of Fe may be incorporated into carbonates, or precipitate as Fe-Mn coatings²⁸. However, while early dolomitisation

in shallow burial environments does not generally cause a significant increase in Fe_{HR} , late stage deep burial dolomitisation may significantly increase Fe_{HR} , but there is no petrographic evidence for deep burial dolomitisation in our samples^{5,29}. Consistent with a recent calibration²⁸, we have limited the application of Fe-speciation to carbonate samples with >0.5wt% Fe_T , which buffers against the impact of non-depositional enrichments in Fe_{HR} ²⁸. In addition, however, we stress that all of our redox interpretations based on Fe speciation in carbonates are entirely consistent with data for interbedded siliciclastics⁵.

Results

In the present study, we measured REY in 259 carbonate rocks of the Nama Group from nine sites across two basins. The succession was deposited ~550-541 Ma broadly coincident with the first appearance of skeletal animals²⁹⁻³¹, as well as trace fossil evidence for motility³² and soft-bodied fossils belonging to the Ediacaran biota³³. Our samples cover a range of palaeo-depths from shallow inner ramp to deeper outer ramp waters, in the Kanies, Omkyk and Hoogland Members of the Kuibis Subgroup, and the Spitzkopf and Feldschuhorn Members of the upper Schwarzrand Subgroup^{5,29} (Fig 2; see Supplementary Information for full details of the geological setting). We focus on the first known skeletal animals, *Cloudina*, a globally distributed eumetazoan of possible cnidarian affinity^{30,34,35}; *Namacalathus*, interpreted as a stem group eumetazoan³⁶ or triploblast lophophorate³⁷ and reported from multiple localities; and *Namapoikia*, an encrusting possible cnidarian or poriferan known only from the Nama Group³¹.

144

145 In the Nama Group, the majority of REY distribution patterns are smooth and show a
146 flat or light REY-depleted shape on shale-normalised plots, positive La anomalies,
147 low total REE concentrations, and superchondritic Y/Ho ratios (>67), all of which
148 indicate preservation and extraction of original seawater signals (see
149 Supplementary Information for a description of all data). Four samples exhibit
150 negative Ce anomalies (<0.9) (Fig 2), consistent with an oxic water column
151 interpretation obtained for these samples from Fe-speciation⁵. Ce anomalies are, as
152 expected, absent from the persistently anoxic and ferruginous deepest water
153 setting⁵ (Fig 2). Significant positive Ce anomalies (1.30 - 2.15) are, however,
154 prevalent in inner ramp sections in both sub-basins (64 samples). In six cases
155 positive Ce anomalies are associated with anoxic ferruginous signals, **and in one**
156 **case, a positive Ce anomaly is associated with a sample that gives a robust oxic**
157 **Fe_{HR}/Fe_T signal. However, for the majority of samples (~90%), Fe_T was <0.5 wt%,**
158 **preventing a robust evaluation of water column redox conditions from Fe speciation**
159 **alone.** In these cases, samples have elevated Mn/Fe ratios (median = 0.39), when
160 compared to samples with no positive Ce anomalies (median = 0.14) and anoxic
161 ferruginous samples (median = 0.10), **which provides an independent constraint on**
162 **water column redox conditions, as discussed below** (Fig 3).

163

164 The regionally widespread positive Ce anomalies across the Zaris and Witputs
165 Basins of the Nama Group imply a surplus of Ce sustained by a rain-down of Mn
166 (oxyhydr)oxides from shallow oxygenated surface waters, and this is supported by

the elevated Mn/Fe ratios of these samples (Fig 3). Redox cycling of Mn (oxyhydr)oxides across the Mn(IV/II) redoxcline would leave ambient waters locally enriched in the Ce released during Mn(IV) reduction (Fig 1). We therefore interpret positive Ce anomalies (>1.3) to indicate intermediate manganous conditions. Where there is an absence of both positive Ce anomalies and any indication of enrichment in Fe (i.e., $Fe_{HR}/Fe_T < 0.22$ or $Fe_T < 0.5$ wt%), we suggest that bottom waters were likely well-oxygenated (which is consistent with Fe_{HR}/Fe_T signals in interbedded siliciclastics⁵), thus preventing the onset of both Fe and Mn reduction. Where data are equivocal (e.g., Fe_{HR}/Fe_T between 0.22-0.38 and no Ce anomaly), we are unable to interpret redox conditions.

The outer ramp was persistently anoxic and ferruginous (Brak section) and animals are absent from these settings⁵ (Fig 4). The deep inner-ramp sections show periods of anoxic ferruginous, manganous and well-oxygenated conditions (Zebra River and Omkyk sections). In these settings, animals are notably absent from ferruginous and manganous waters, whereas well-oxygenated waters support abundant skeletal animals, up to 35 mm in diameter, and adjacent localities show trace fossil evidence for motility³². The shallowest inner ramp sections show high-frequency temporal fluctuations between anoxic ferruginous, manganous and well-oxygenated conditions (Zwartmodder, Arasab and Grens sections), as might be expected due to fluctuations in the depth of the chemocline (Fig 4). At Zwartmodder, skeletal animals are present in thin beds⁵, but there is only one skeletal horizon at Grens, and no animal fossils at Arasab.

In contrast to these ecologies, the Driedoornvlagte pinnacle reef grew within a transgressive systems tract in a mid-ramp position, which was persistently well-oxygenated and hosts some very large skeletal animals^{5,31} (up to 1 m) and complex reef-building ecologies³⁰. In the younger Schwarstrand Subgroup, which extends close to the Ediacaran-Cambrian Boundary (~547-541 Ma), there is evidence for persistent well-oxygenated conditions⁵ and mid-ramp Pinnacle Reefs host mixed communities of large and small skeletal animals. At Swartpunt, abundant burrows and soft-bodied biota occur in siliciclastic horizons, where Fe speciation indicates oxic conditions⁵, while small *in-situ* skeletal animals are found in carbonate rocks throughout the succession⁵.

Discussion

Our geochemical and palaeontological data demonstrate a striking relationship between the precise redox condition of the water column and the presence and abundance of evidence for animal life. Constraints from the modern open ocean suggest that dissolved Mn(II), and therefore Ce(III), can start to build up in low concentrations in oxic waters with dissolved O₂ <100 µM¹⁷. However, manganous conditions, whereby Mn becomes the dominant redox buffer, are achieved at lower oxygen concentrations. Reduced Mn can remain stable in the presence of up to 10 µM O₂^{16,38}, although Mn oxidation has been reported locally at lower O₂ concentrations where oxidation is catalysed by enzymatic processes³⁹. Thus, active Mn cycling can occur in anoxic waters, but is commonly documented in partially oxic

waters with at least 10 $\mu\text{M O}_2$ (and up to 100 $\mu\text{M O}_2$; Fig 1)^{16,38,40,41}, which represents significant oxygen depletion in comparison to modern fully oxygenated surface waters (~250 $\mu\text{M O}_2$). The reduction potential for Ce is higher than that for Mn, and so the 10 $\mu\text{M O}_2$ constraint for manganous waters may represent a lower limit on Ce cycling, as sufficient O_2 to oxidise both Ce and Mn is required for the formation of Ce anomalies.

Our multi-proxy approach allows us to distinguish between fully anoxic and intermediate waters, which contained low but significant amounts of oxygen. Where Fe-speciation in Ce enriched samples gives a robust anoxic signal ($\text{Fe}_{\text{HR}}/\text{Fe}_{\text{T}} > 0.38$), Mn reduction may have persisted, but conditions must have been fully anoxic. However, the majority of samples interpreted to be manganous have insufficient Fe_{T} for Fe-speciation (with 85% of these falling below 0.25 wt% Fe_{T} , and 35% falling below 0.1 wt% Fe_{T}). Even very low oxygen concentrations (nM) are sufficient to prevent highly reactive Fe enrichments, and so the low Fe_{T} in shallower environments across the Nama Group may be indicative of oxic conditions⁴², and this is supported by persistent oxic $\text{Fe}_{\text{HR}}/\text{Fe}_{\text{T}}$ ratios obtained from interbedded siliciclastics in some sections⁵. We therefore suggest that the manganous zone occurred between well-oxygenated surface waters and deeper anoxic, ferruginous waters, commonly overlapping with low but significant concentrations of oxygen (at least ~10 μM) (Fig 1).

235 Oxygen exerts an important control on ecosystem structure in modern
236 environments, whereby low oxygen environments are inhabited by smaller animals
237 often lacking skeletons and forming low diversity communities with simple food
238 webs⁴³. In general, skeletons are absent from modern oxygen minimum zones when
239 O₂ drops below 13 μM, and large animals are often absent below 45 μM [refs ^{44,45}].
240 However, the importance of oxygen in supporting early animal ecosystems as they
241 became increasingly complex in form, metabolic demand, and behaviour through
242 the Ediacaran Period is currently unresolved^{2,3,5,11-13}. In the Nama Group the
243 majority of small skeletal animals (>75%), and all evidence for large skeletal
244 animals, motility, soft-bodied biota and complex ecologies³⁰, are found in sediments
245 deposited from well-oxygenated waters (Fig 4). The identification of low-oxygen,
246 manganoous water column conditions thus provides a compelling explanation for the
247 general absence of biota in these settings, and implies that poorly oxygenated
248 conditions were insufficient to meet the relatively high oxygen requirements of
249 these early skeletal animals^{5,30,32,33}. If we take an upper O₂ limit for Mn and Ce
250 reduction of 10 μM O₂, this suggests that Mn-enriched waters could theoretically
251 support small, soft-bodied animals, such as sponges¹⁵. In contrast, the absence of
252 skeletal animals in Mn-enriched waters is consistent with the high energetic cost of
253 skeletonisation. Possible biomarkers for sponge animals appear in the fossil record
254 at >635 Ma [Ref 46], but it is possible that the availability of well-oxygenated
255 habitats was necessary to support the later appearance of skeletonisation, at ~550
256 Ma. However, it is also **unlikely** that reaching an oxygenation threshold alone is
257 sufficient to explain the appearance of skeletons⁴⁷, and many have argued that the


trigger for the rise of skeletonisation may have been ecological, such as the rise of predation^{36,48}.

Our approach highlights that intermediate redox conditions were probably widespread in the Ediacaran ocean, but have not previously been appreciated due to the inability of most commonly used proxies to identify such conditions. Our data suggest that low oxygen water column conditions were insufficient to support early skeletal and reef-building animals, and so the extent of suitable habitat space may have been less than previously identified. The widespread radiation of skeletal animals during the subsequent Cambrian explosion may have been facilitated by a global rise in the extent of habitable, oxygenated seafloor⁷, alongside other genetic and ecological factors. Our data therefore yield new insight into the debate on the role of oxygen in early animal evolution, suggesting that well-oxygenated waters were necessary to support the appearance of the skeletal animals and complex ecologies that are typical of the terminal Neoproterozoic.

273

- 274 1. Poulton, S. W. & Canfield, D. E. Ferruginous Conditions: A Dominant Feature of
275 the Ocean through Earth's History. *ELEMENTS* **7**, 107–112 (2011).
- 276 2. Johnston, D. T. *et al.* Late Ediacaran redox stability and metazoan evolution.
277 *Earth and Planetary Science Letters* **335**, 25–35 (2012).
- 278 3. Planavsky, N. J. *et al.* Low Mid-Proterozoic atmospheric oxygen levels and the
279 delayed rise of animals. *Science* **346**, 635–638 (2014).
- 280 4. Canfield, D. E., Poulton, S. W. & Narbonne, G. M. Late-Neoproterozoic Deep-Ocean
281 Oxygenation and the Rise of Animal Life. *Science* **315**, 92 –95 (2007).
- 282 5. Wood, R. A. *et al.* Dynamic redox conditions control late Ediacaran ecosystems in
283 the Nama Group, Namibia. *Precambrian Research* **261**, 252–271 (2015).
- 284 6. Scott, C. *et al.* Tracing the stepwise oxygenation of the Proterozoic ocean. *Nature*
285 **452**, 456–459 (2008).
- 286 7. Chen, X. *et al.* Rise to modern levels of ocean oxygenation coincided with the
287 Cambrian radiation of animals. *Nat Commun* **6**, (2015).
- 288 8. Poulton, S. W., Fralick, P. W. & Canfield, D. E. Spatial variability in oceanic redox
289 structure 1.8 billion years ago. *Nature Geoscience* **3**, 486–490 (2010).
- 290 9. Guilbaud, R., Poulton, S. W., Butterfield, N. J., Zhu, M. & Shields-Zhou, G. A. A
291 global transition to ferruginous conditions in the early Neoproterozoic oceans.
292 *Nature Geosci* **8**, 466–470 (2015).
- 293 10. Sperling, E. A. *et al.* Statistical analysis of iron geochemical data suggests limited
294 late Proterozoic oxygenation. *Nature* **523**, 451–454 (2015).

- 295 11. Nursall, J. Oxygen as a prerequisite to the origin of the Metazoa. *Nature* **183**,
296 1170–1172 (1959).
- 297 12. Lenton, T. M., Boyle, R. A., Poulton, S. W., Shields-Zhou, G. A. & Butterfield, N. J.
298 Co-evolution of eukaryotes and ocean oxygenation in the Neoproterozoic era.
299 *Nature Geoscience* (2014).
- 300 13. Butterfield, N. J. Oxygen, animals and oceanic ventilation: an alternative view.
301 *Geobiology* **7**, 1–7 (2009).
- 302 14. Sperling, E. A. *et al.* Oxygen, facies, and secular controls on the appearance of
303 Cryogenian and Ediacaran body and trace fossils in the Mackenzie Mountains of
304 northwestern Canada. *Geological Society of America Bulletin* B31329.1 (2015).
- 305 15. Mills, D. B. *et al.* Oxygen requirements of the earliest animals. *Proceedings of the*
306 *National Academy of Sciences* **111**, 4168–4172 (2014).
- 307 16. Johnson, K. S. *et al.* Manganese Flux from Continental Margin Sediments in a
308 Transect Through the Oxygen Minimum. *Science* **257**, 1242–1245 (1992).
- 309 17. Klinkhammer, G. P. & Bender, M. L. The distribution of manganese in the Pacific
310 Ocean. *Earth and Planetary Science Letters* **46**, 361–384 (1980).
- 311 18. Sholkovitz, E. R., Landing, W. M. & Lewis, B. L. Ocean particle chemistry: The
312 fractionation of rare earth elements between suspended particles and seawater.
313 *Geochimica et Cosmochimica Acta* **58**, 1567–1579 (1994).
- 314 19. Ohta, A. & Kawabe, I. REE (III) adsorption onto Mn dioxide (MnO₂) and Fe
315 oxyhydroxide: Ce (III) oxidation by MnO₂. *Geochimica et Cosmochimica Acta* **65**,
316 695–703 (2001).

- 317 20. De Carlo, E. H. & Green, W. J. Rare earth elements in the water column of Lake
318 Vanda, McMurdo Dry Valleys, Antarctica. *Geochimica et Cosmochimica Acta* **66**,
319 1323–1333 (2002).
- 320 21. de Baar, H. J. ., German, C. R., Elderfield, H. & van Gaans, P. Rare earth element
321 distributions in anoxic waters of the Cariaco Trench. *Geochimica et*
322 *Cosmochimica Acta* **52**, 1203–1219 (1988).
- 323 22. Bau, M., Möller, P. & Dulski, P. Yttrium and lanthanides in eastern Mediterranean
324 seawater and their fractionation during redox-cycling. *Marine Chemistry* **56**,
325 123–131 (1997).
- 326 23. Webb, G. E. & Kamber, B. S. Rare earth elements in Holocene reefal microbialites:
327 a new shallow seawater proxy. *Geochimica et Cosmochimica Acta* **64**, 1557–1565
328 (2000).
- 329 24. Banner, J. L., Hanson, G. N. & Meyers, W. J. Rare Earth Element and Nd Isotopic
330 Variations in Regionally Extensive Dolomites From the Burlington-Keokuk
331 Formation (Mississippian): Implications for Reef Mobility During Carbonate
332 Diagenesis. *Journal of Sedimentary Research* **58**, (1988).
- 333 25. Tostevin, R. *et al.* Effective use of Ce anomalies as a redox proxy in carbonate
334 dominated marine settings. (accepted). 
- 335 26. Himmler, T., Bach, W., Bohrmann, G. & Peckmann, J. Rare earth elements in
336 authigenic methane-seep carbonates as tracers for fluid composition during
337 early diagenesis. *Chemical Geology* **277**, 126–136 (2010).
- 338 27. Raiswell, R. & Canfield, D. E. Sources of iron for pyrite formation in marine
339 sediments. *American Journal of Science* **298**, 219–245 (1998).

- 340 28. Clarkson, M. O., Poulton, S. W., Guilbaud, R. & Wood, R. A. Assessing the utility of
341 Fe/Al and Fe-speciation to record water column redox conditions in carbonate-
342 rich sediments. *Chemical Geology* **382**, 111-122 (2014).
- 343 29. Grotzinger, J. P., Watters, W. A. & Knoll, A. H. Calcified metazoans in thrombolite-
344 stromatolite reefs of the terminal Proterozoic Nama Group, Namibia.
345 *Paleobiology* **26**, 334 –359 (2000).
- 346 30. Penny, A. *et al.* Ediacaran metazoan reefs from the Nama Group, Namibia. *Science*
347 **344**, 1504–1506 (2014).
- 348 31. Wood, R. A., Grotzinger, J. P. & Dickson, J. A. D. Proterozoic Modular
349 Biomineralized Metazoan from the Nama Group, Namibia. *Science* **296**, 2383 –
350 2386 (2002).
- 351 32. Macdonald, F. A., Pruss, S. B. & Strauss, J. V. Trace fossils with spreiten from the
352 late Ediacaran Nama Group, Namibia: complex feeding patterns five million years
353 before the Precambrian–Cambrian boundary. *Journal of Paleontology* **88**, 299–
354 308 (2014).
- 355 33. Hall, M. *et al.* Stratigraphy, palaeontology and geochemistry of the late
356 Neoproterozoic Aar Member, southwest Namibia: Reflecting environmental
357 controls on Ediacara fossil preservation during the terminal Proterozoic in
358 African Gondwana. *Precambrian Research* **238**, 214–232 (2013).
- 359 34. Morris, S. C., Mattes, B. & Chen, M. The early skeletal organism Cloudina: new
360 occurrences from Oman and possibly China. *American Journal of Science* 245–
361 260 (1990).

- 362 35. Warren, L. V. *et al.* Corumbella and in situ Cloudina in association with
363 thrombolites in the Ediacaran Itapucumi Group, Paraguay. *Terra Nova* **23**, 382–
364 389 (2011).
- 365 36. Wood, R., A. Paleoecology of the earliest skeletal metazoan communities:
366 Implications for early biomineralization. *Earth-Science Reviews* **106**, 184–190
367 (2011).
- 368 37. Zhuravlev, A. Y., Wood, R. A. & Penny, A. M. Ediacaran skeletal metazoan
369 interpreted as a lophophorate. *Proc. R. Soc. B* **282**, 20151860 (2015).
- 370 38. German, C. R. & Elderfield, H. Rare earth elements in the NW Indian Ocean.
371 *Geochimica et Cosmochimica Acta* **54**, 1929–1940 (1990).
- 372 39. Clement, B. G., Luther, G. W., III & Tebo, B. M. Rapid, oxygen-dependent microbial
373 Mn(II) oxidation kinetics at sub-micromolar oxygen concentrations in the Black
374 Sea suboxic zone. *Geochimica et Cosmochimica Acta* **73**, 1878–1889 (2009).
- 375 40. Saager, P. M., De Baar, H. J. W. & Burkill, P. H. Manganese and iron in Indian
376 Ocean waters. *Geochimica et Cosmochimica Acta* **53**, 2259–2267 (1989).
- 377 41. Trefry, J. H., Presley, B. J., Keeney-Kennicutt, W. L. & Trocine, R. P. Distribution
378 and chemistry of manganese, iron, and suspended particulates in Orca Basin.
379 *Geo-Marine Letters* **4**, 125–130 (1984).
- 380 42. Raiswell, R. & Anderson, T. F. Reactive iron enrichment in sediments deposited
381 beneath euxinic bottom waters: constraints on supply by shelf recycling.
382 *Geological Society, London, Special Publications* **248**, 179–194 (2005).

- 383 43. Gibson, R. & Atkinson, R. Oxygen minimum zone benthos: adaptation and
384 community response to hypoxia. *Oceanography and Marine Biology: An Annual*
385 *Review* **41**, 1–45 (2003).
- 386 44. Levin, L. A., Gage, J. D., Martin, C. & Lamont, P. A. Macrobenthic community
387 structure within and beneath the oxygen minimum zone, NW Arabian Sea. *Deep*
388 *Sea Research* **47**, 189–226 (2000).
- 389 45. Savrda, C. E. & Bottjer, D. J. Oxygen-related biofacies in marine strata: an
390 overview and update. *Geological Society, London, Special Publications* **58**, 201–
391 219 (1991).
- 392 46. Love, G. D. *et al.* Fossil steroids record the appearance of Demospongiae during
393 the Cryogenian period. *Nature* **457**, 718–721 (2009).
- 394 47. Zhang, S. *et al.* Sufficient oxygen for animal respiration 1,400 million years ago.
395 *PNAS* **113**, 1731–1736 (2016).
- 396 48. Knoll, A. H. Biomineralization and Evolutionary History. *Reviews in Mineralogy*
397 *and Geochemistry* **54**, 329–356 (2003).
- 398 49. Turekian, K. K. & Wedepohl, K. H. Distribution of the Elements in Some Major
399 Units of the Earth's Crust. *Geological Society of America Bulletin* **72**, 175–192
400 (1961).
- 401 50. Grotzinger, J. & Miller, R. *The Nama Group*. **2**, (Geological Society of Namibia,
402 2008).
- 403 51. Bau, M. & Dulski, P. Distribution of yttrium and rare-earth elements in the Penge
404 and Kuruman iron-formations, Transvaal Supergroup, South Africa. *Precambrian*
405 *Research* **79**, 37–55 (1996).

52. Sunda, W. G. & Huntsman, S. A. Effect of sunlight on redox cycles of manganese in the southwestern Sargasso Sea. *Deep Sea Research Part A. Oceanographic Research Papers* **35**, 1297–1317 (1988).
53. Bau, M., Koschinsky, A., Dulski, P. & Hein, J. R. Comparison of the partitioning behaviours of yttrium, rare earth elements, and titanium between hydrogenetic marine ferromanganese crusts and seawater. *Geochimica et Cosmochimica Acta* **60**, 1709–1725 (1996).
54. Byrne, R. H., Liu, X. & Schijf, J. The influence of phosphate coprecipitation on rare earth distributions in natural waters. *Geochimica et Cosmochimica Acta* **60**, 3341–3346 (1996).
55. German, C. R. & Elderfield, H. Application of the Ce anomaly as a paleoredox indicator: The ground rules. *Paleoceanography* **5**, 823–833 (1990).
56. Shields, G. & Webb, G. Has the REE Composition of Seawater Changed Over Geological Time? **204.1**, 103–107 (2004).
57. Poulton, S. W. & Canfield, D. E. Development of a sequential extraction procedure for iron: implications for iron partitioning in continentally derived particulates. *Chemical Geology* **214**, 209–221 (2005).
58. Canfield, D. E., Raiswell, R., Westrich, J. T., Reaves, C. M. & Berner, R. A. The use of chromium reduction in the analysis of reduced inorganic sulfur in sediments and shales. *Chemical Geology* **54**, 149–155 (1986).

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Author contributions

RAW, ARP, KHH, RT, AMP, FB, and AC collected the samples. MOC, RT, AMP and FB prepared the samples. RT conceived the project and analysed the samples. RT interpreted the Ce anomaly data, after discussions with GAS, TH, RG and SWP. RT wrote the paper, with significant input from GAS, SWP, RAW, and RG.


Figure 1



Schematic representation of redox zones and associated geochemical signals in the Zaris Basin, Nama Group during a highstand systems tract. Positive Ce anomalies form as Mn (oxyhydr)oxides dissolve in the manganous zone, and Fe enrichments form under anoxic ferruginous conditions. 10 μM is an estimate of maximum O_2 concentrations in the manganous zone, but overlying well-oxygenated waters may have contained higher O_2 concentrations. Representative REY patterns, including positive Ce anomalies (magnitude in brackets), are shown for the Omkyk section in

the Nama Group, alongside manganoan zones from two modern environments^{20,22}
(modern water column data plotted as $[REY] \times 10^6$ to be comparable with
sedimentary $[REY]$).

Figure

The location of nine sections within the Kuibis and Schwarzrand Subgroups of the
Nama Group is shown on a simplified geological map of Namibia (top left), as well as
on a schematic cross section indicating average relative water depth (bottom). Fe_T
data for each location is shown for carbonate and siliciclastic rocks, colour coded for
redox interpretation based on Fe_{HR}/Fe_T and Fe_T^{5+} , alongside Ce_{SN}/Ce_{SN}^* data,
screened for carbonate rocks showing seawater REY patterns (e.g. molar $Y/Ho > 67$).
Blue Fe_T data indicate where Fe speciation would predict oxic conditions⁵, and
positive Ce anomalies indicate where oxic waters are interpreted  to have been
manganoan. The presence of *in situ* biota is noted by grey lines⁵.

Figure


Mn/Fe ratios for samples identified as manganoan (positive Ce/Ce^* and low Fe_T or
oxic Fe-speciation signals), ferruginous (anoxic Fe-speciation signals) and oxic (oxic
Fe-speciation signals, no positive Ce anomaly). Mn/Fe is enriched in manganoan
samples compared with global carbonate (0.29)[Ref 49]. Bars represent median
values. Red o  nes indicate dolomitised samples.

Figure 4

A comprehensive redox interpretation is shown for each of the nine localities in the Nama Group, determined using combined Fe and Ce signals (see inset table). *In situ* fossils (grey lines), and local ecologies⁵ and general ecology from the literature³⁰ are shown alongside local water column redox conditions. The bar chart plots the frequency that different biota are found in each redox zone⁵. Large skeletal fossils and burrows are found exclusively in well-oxygenated settings, and small skeletal fossils are largely restricted to well-oxygenated conditions, but may occur where conditions were only fleetingly oxid.

Figure 1

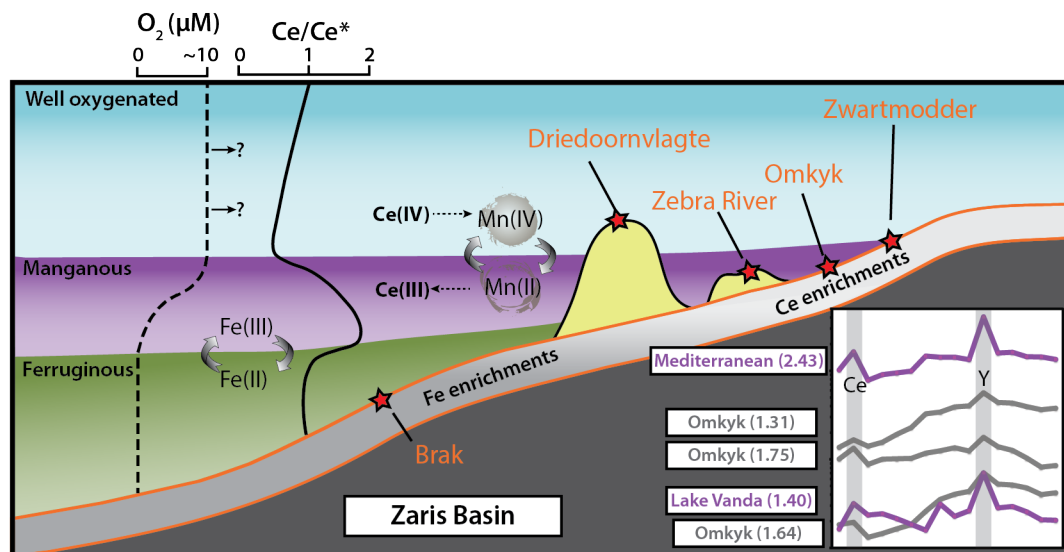


Figure 2

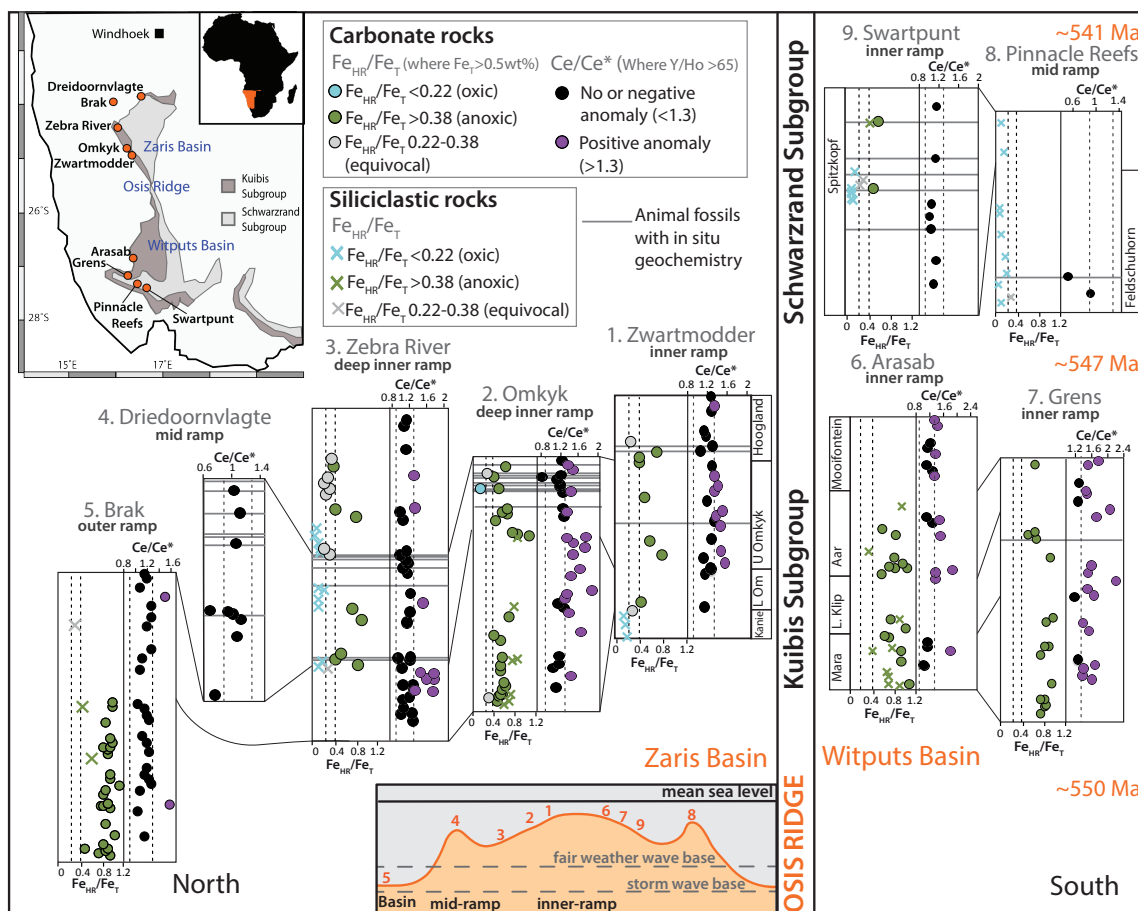
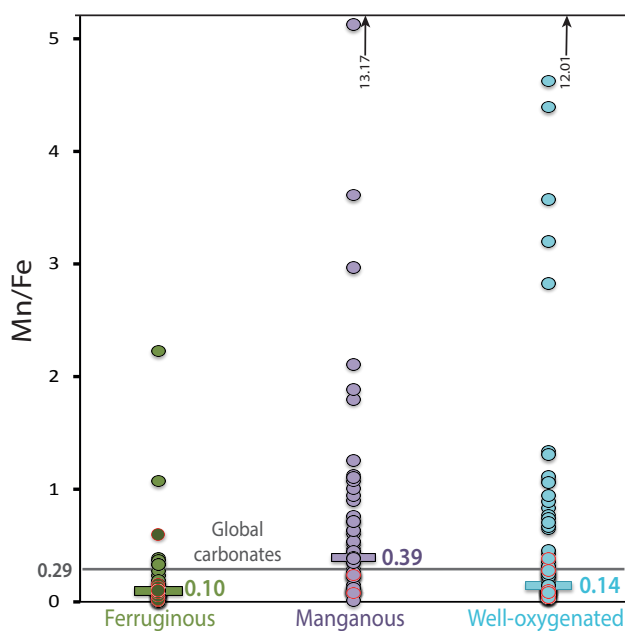
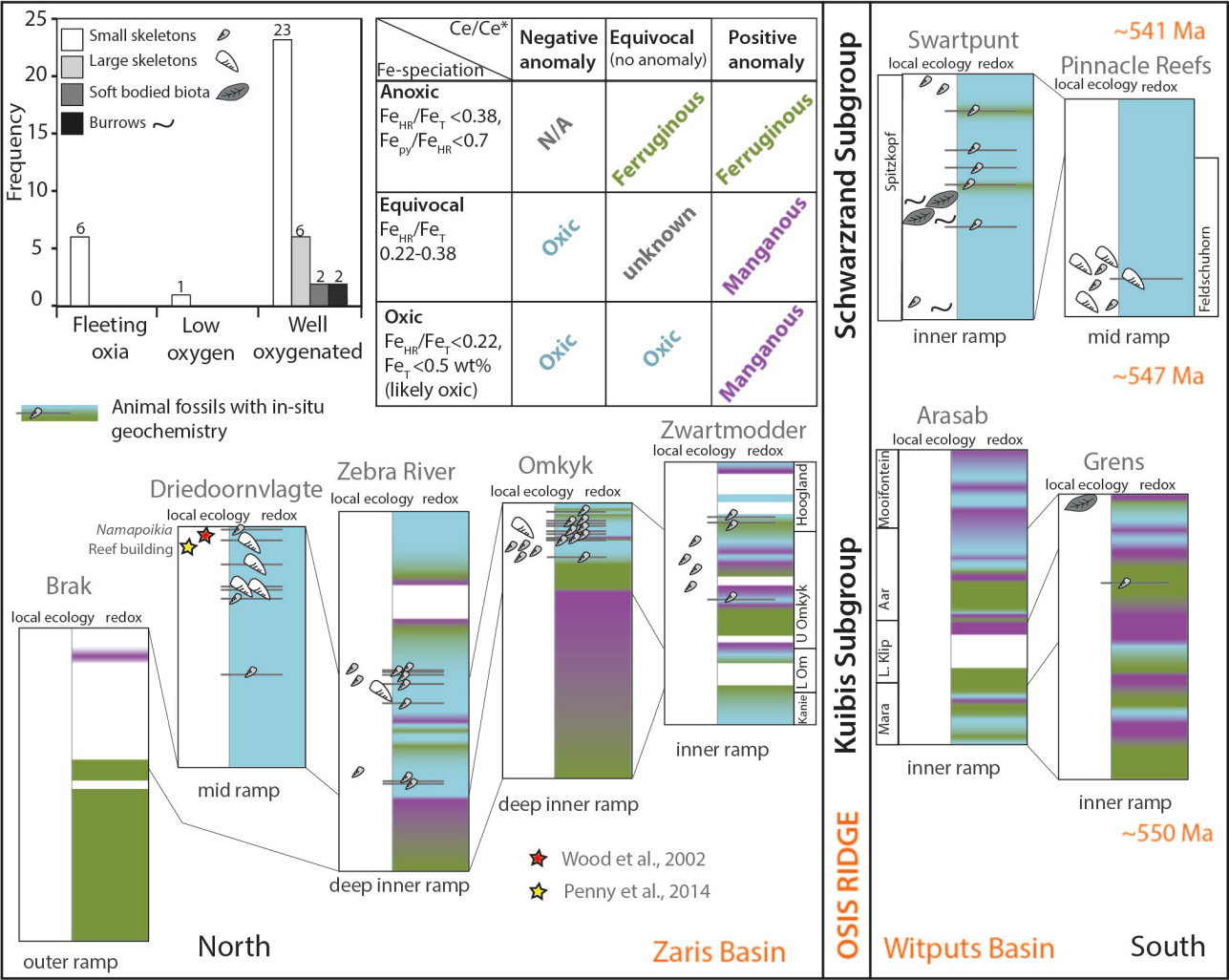


Figure 3




492 **Figure 4**



493
494
495

496 **Methods**

497 Samples from nine shelf-to-basin sections within the Zaris and Witputs basins of the
498 Nama Group encompass a range of palaeo-depths from outer- to inner-ramp
499 settings. Stratigraphic correlations are well-established based on sequence
500 boundaries and ash beds^{5,29}. Unweathered samples were selected, and powdered or
501 drilled avoiding alteration, veins or weathered edges. **For Zebra River section,**
502 **powders were drilled from thin section counterparts, targeting fine grained**
503 **cements.** Carbonate rocks in the Nama Group are very pure, but they have all
504 undergone pervasive recrystallisation. Less than 15% of the samples in this study
505 are dolomitised, and there is no petrographic evidence for deep burial
506 dolomitisation in the Nama Group^{5,50}. Most carbonate rocks are associated with low
507 Mn/Sr ratios and heavy $\delta^{18}\text{O}$ (see Supplement  information). The presence of
508 different forms of skeletal biota, soft-bodied biota, trace fossils are reported for
509 precise horizons where geochemical analyses have been performed⁵, indicated by
510 grey lines on Figs 2 and 3. General local ecology, supported by additional
511 information from the literature, is also marked, without associated grey lines. Our
512 sampling focused on carbonates, and hence skeletal fossils are over-represented
513 compared with soft-bodied biota and trace fossils. We define 'large' skeletal animals
514 as >10 mm in any dimension, which includes *Cloudina hartmannae*, some
515 *Namacalathus*, and *Namapoikia*.

516 **Rare earth elements**

Rare earth elements and yttrium (REY) have a predictable distribution pattern in seawater, and non-biological carbonate rocks should preserve local water column REY at the sediment-water interface²³. Ce anomalies develop progressively, but cut-off values are established to define negative and positive anomalies. We define a negative anomaly as $Ce_{SN}/Ce_{SN}^* < 0.9$, consistent with previous work⁵¹. A positive anomaly, using the same reference frame, would be defined as $Ce_{SN}/Ce_{SN}^* > 1.1$. But since positive anomalies are not previously described from carbonate sediments, we cautiously use a higher cut off, $Ce_{SN}/Ce_{SN}^* > 1.3$, to ensure any positive anomalies are environmentally significant with respect to positive anomalies recorded from some modern manganoous waters (1.21-2.43) (see supplementary information for discussion of Ce_{SN}/Ce_{SN}^* cut-offs). While positive or negative Ce anomalies in carbonate rocks likely represent seawater redox conditions, the absence of any Ce anomaly (0.9-1.3) is somewhat equivocal, and could result from anoxic water column conditions, or overprinting of any Ce anomaly during diagenesis or leaching²⁶. Alternately, Ce **anomaly formation may be disrupted** in surface waters because of wind-blown dust or photo-reduction of Mn oxides⁵². Fe (oxyhydr)oxides may also be REY carriers, but do not contain the clear Ce enrichments observed in Mn (oxyhydr)oxides (see Supplementary **Information for full discussion of REY carriers**).

Diagenetic phosphates, Fe and Mn (oxyhydr)oxides, organic matter and clays can potentially affect the REY signatures of authigenic sedimentary rocks if they are partially dissolved during the leaching process⁵³⁻⁵⁶. Care has been taken to partially

leach samples, to isolate the carbonate phase without leaving excess acid, which may leach contaminant phases (see Ref 26 for detailed discussion of methodology).

Powdered calcite samples were cleaned in Milli-Q water and pre-leached in 2% nitric acid to remove adsorbed and easily exchangeable ions associated with clay minerals. The remaining sample was partially leached, also in 2% (w/v) nitric acid, to avoid contributions from contaminant phases such as oxides and clays²⁶. The supernatant was removed from contact with the remaining residue, diluted with 2% nitric acid and analysed via inductively coupled plasma mass spectrometry in the Cross-Faculty Elemental Analysis Facility, University College London. This leaching method has been designed to extract the carbonate bound REY pool without contributions from (oxyhydr)oxides or clays²⁶. These same leachates were also analysed for major element concentrations (Mg, Fe, Mn, Al and Sr) via inductively coupled plasma optical emission spectrometry. Oxide interference was monitored using the formation rate of Ce oxide, and the formation of 2+ ions was monitored using Ba²⁺. All REY concentrations were normalised to post-Archean Australian Shale (PAAS). Ce anomalies were calculated using equation (1).

Standard solutions analysed after every ten samples were within 5% of known concentrations. Replicate analyses on the ICP-MS give a relative standard deviation <5% for most trace elements, with a larger standard deviation for the heavy REE that sometimes have non-normalised concentrations <0.5 ppb. Carbonate standard material CRM 1c was prepared using the same leaching procedure as the samples, and repeat analyses give a relative standard deviation <5% for individual REY

concentrations, and calculated Ce anomalies (average=0.80) give a relative standard deviation <3%.

Mn/Sr ratios are <1 for the majority (97%) of samples, and $\delta^{18}\text{O}_{\text{carb}}$ is >-10‰, indicating minimal open-system elemental and isotopic exchange during diagenesis, and excluding deep burial dolomitisation (Fig S3). Ce anomaly data are only presented for carbonates that preserve seawater REY features (smooth patterns with molar Y/Ho>67)^{23,26} indicating they originate from the carbonate portion of the whole rock, without contributions from detrital or oxide phases. For samples with Y/Ho>67, 85% also have ΣREE <2 ppm, and all have ΣREE <10 ppm. La anomalies, and small positive Eu and Gd enrichments, are prevalent in samples with Y/Ho>67. Positive Ce anomalies are associated with low Mn/Sr ratios (<1) and low Al, Zr, Ti, Fe and Mn contents in the leachate (<0.2 wt% for Fe, and <500 ppm for Mn), indicating minimal contamination due to diagenetic exchange, leaching of clays or Fe-Mn (oxyhydr)oxide phases (see Supplementary Information for a full assessment of data quality).

Fe speciation

The Fe speciation method quantifies Fe that is (bio)geochemically available in surficial environments (termed highly reactive Fe; Fe_{HR}) relative to total Fe (Fe_{T}). Mobilisation and subsequent precipitation of Fe in anoxic water column settings results in Fe_{HR} enrichments in the underlying sediment. The nature of anoxia (i.e. sulphide-rich or Fe-containing) is determined by the extent of sulphidation of the

highly reactive Fe pool¹. Fe speciation data for carbonate rock samples discussed here, and accompanying interbedded siliciclastic rocks, come from previously published data⁵. The Fe-speciation technique was performed using well established sequential extraction schemes²⁹. The method targets operationally defined Fe pools, including carbonate-associated-Fe (Fe_{Carb}), ferric oxides (Fe_{Ox}), magnetite (Fe_{Mag}), pyrite Fe (Fe_{Py}) and Fe_{T} . Fe_{HR} is defined as the sum of Fe_{carb} (extracted with Na-acetate at pH 4.5 and 50°C for 48h), Fe_{ox} (extracted via Na-dithionite at pH 4.8 for 2h), Fe_{mag} (extracted with ammonium oxalate for 6h) and Fe_{py} (calculated from the mass of sulphide extracted during CrCl_2 distillation). Fe_{T} extractions were performed on ashed samples (8 h at 550°C) using $\text{HNO}_3\text{--HF--H}_3\text{BO}_3\text{--HClO}_4$. All Fe concentrations were measured via atomic absorption spectrometry and replicate extractions gave a relative standard deviation of <4% for all steps, leading to <8% for calculated Fe_{HR} (see Supplementary Information for full discussion of data quality). Fe_{Py} was calculated from the wt% of sulphide extracted as Ag_2S using hot Cr(II)Cl_2 distillation⁵⁸. A boiling HCl distillation before the Cr(II)Cl_2 distillation ruled out the potential presence of acid volatile sulfides in our samples. Pyrite extractions give reproducibility for Fe_{py} of 0.005 wt%, confirming high precision for this method. Analysis of a certified reference material (PACS-2, $\text{Fe}_{\text{T}} = 4.09 \pm 0.07$ wt%, $n = 4$; certified value = 4.09 ± 0.06 wt%) confirms that our method is accurate. Replicate analyses ($n = 6$) gave a precision of ± 0.06 wt% for Fe_{T} , and a relative standard deviation of <5% for the $\text{Fe}_{\text{HR}}/\text{Fe}_{\text{T}}$ ratio.

609 Calibration in modern and ancient marine environments suggests that $\text{Fe}_{\text{HR}}/\text{Fe}_{\text{T}}$
610 <0.22 indicates deposition under oxic water column conditions, while $\text{Fe}_{\text{HR}}/\text{Fe}_{\text{T}}$
611 >0.38 indicates anoxic conditions¹. Ratios between 0.22–0.38 are considered
612 equivocal, and may represent either oxic or anoxic depositional conditions. For
613 sediments identified as anoxic, $\text{Fe}_{\text{py}}/\text{Fe}_{\text{HR}} >0.8$ is diagnostic for euxinic conditions
614 and $\text{Fe}_{\text{py}}/\text{Fe}_{\text{HR}} <0.7$ defines ferruginous deposition¹. This also applies to carbonate-
615 rich sediments that have not undergone late stage dolomitisation²⁸, on the condition
616 that $\text{Fe}_{\text{T}} >0.5\text{wt}\%$. Where Fe_{T} is very low ($<0.5\text{wt}\%$), this may indicate deposition
617 under oxic conditions²⁸. However, wherever possible we consider together data
618 obtained from siliciclastic horizons interbedded with and/or associated with
619 carbonate rocks contained within the same m- to dm-scale depositional cycle. Fe-
620 speciation in carbonate and siliciclastic rocks gives consistent signals in the Nama
621 Group (Figure 2), supporting the application of this proxy in carbonate rocks^{5,28}